

Characterization of the α -Glucuronyl C5-epimerase Involved in the Biosynthesis of Heparin and Heparan Sulfate*

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The murine gene for the glucuronyl CS-epimerase involved in heparan sulfate biosynthesis was cloned, using a previously isolated bovine lung cDNA fragment (Li, J.-P., Wagner-McWhirter, A., Kjellén, L., Palgi, J., Jalkanen, M., and Lindahl, U. (1997) *J. Biol. Chem.* 272, 28158–28163) as probe. The ~11-kilobase pair mouse gene contains 3 exons from the first ATG to stop codon and is localized to chromosome 9. Southern analysis of the genomic DNA and chromosome mapping suggested the occurrence of a single epimerase gene. Based on the genomic sequence, a mouse liver cDNA was isolated that encodes a 618-amino acid residue protein, thus extending by 174 N-terminal residues the sequence deduced from the (incomplete) bovine cDNA. Comparison of murine, bovine, and human epimerase cDNA structures indicated 96–99% identity at the amino acid level. A cDNA identical to the mouse liver species was demonstrated in mouse mast cells committed to heparin biosynthesis. These findings suggest that the iduronic acid residues in heparin and heparan sulfate, despite different structural contexts, are generated by the same CS-epimerase enzyme. The catalytic activity of the recombinant full-length mouse liver epimerase, expressed in insect cells, was found to be >2 orders of magnitude higher than that of the previously cloned, smaller bovine recombinant protein. The ~32-kDa, similarly highly active, enzyme originally purified from bovine liver (Campbell, P., Hammarstrom, H. H., Sandback, D., Rodén, I., Lindahl, U., and Li, J.-P. (1994) *J. Biol. Chem.* 269, 26953–26958) was found to be associated with an ~22-kDa peptide generated by a single proteolytic cleavage of the full-sized protein.

Heparin and heparan sulfate (HS)[†] are both linear, sulfated glycosaminoglycans based on a common carbohydrate back-

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[†] The nucleotide sequence(s) reported in this paper has been submitted to the GenBank™/EMBL Data Bank with accession number(s) AF330049 and AF0000527.

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[‡] The abbreviations used are: HS, heparan sulfate; IdUA, α -iduronic acid; IdoUA, β -iduronic acid; GAG, glycosaminoglycan; FITC, fluorescein isothiocyanate; DAPI, 4',6-diamidino-2-phenylindole; α Mann, 2,3-anhydrofructose (formed by reduction of terminal 2,3-anhydrofructose residues with NaBH₄); PAGE, polyacrylamide gel

bone of alternating α -glucosamine (GlcNAc) and hexuronic acid (α -glucuronic (GlcUA) or β -iduronic acid (IdoUA)) units. Both polymer types are synthesized as proteoglycans. Heparin occurs in connective tissue-type mast cells, whereas HS has a ubiquitous distribution and is produced by almost all mammalian cells. The biosynthesis of heparin and HS follows a common pathway that involves: (i) formation of a GlcUA-Gal-Gal-Xyl-Ser carbohydrate-protein linkage region; (ii) assembly of repeating -GlcUA-GlcNAc_n-disaccharide units; and (iii) modification of the [GlcUA-GlcNAc]_n polymers (1, 2). The latter process includes N -deacetylation and N -sulfation of GlcNAc units, C5-epimerization of GlcUA to IdoUA units, and O-sulfation at various sites of the sugar residues. Heparin is highly modified, heavily N - and O-sulfated and with more IdoUA than GlcUA units. Generally, a predominant proportion of heparin chains consists of trisulfated -IdoUA(2-O SO_3^-)-GlcNSO₃(6-O SO_3^-)-units. By comparison, HS has a more variable and heterogeneous structure, with highly modified, less modified, and unmodified sequences arranged in domain-type fashion (3, 4). The IdoUA units occur in essentially two types of domains, composed of contiguous N -sulfated, and of alternating N -sulfated and N -acetylated disaccharide units, respectively (5). The sequence variability of the chains is believed to reflect the functional role of HS glycosaminoglycans (GAGs) in specific interactions with different proteins (1–4, 6). IdoUA residues generally appear to promote protein binding due to their conformational flexibility and have been identified as invariable constituents of protein-binding HS domains (4). The reaction catalyzed by the CS-epimerase therefore is crucial for many biological functions of heparin and HS.

The mechanisms in control of the structural diversity of heparin and, in particular, of HS are only partly understood but clearly rely on the substrate specificities of the enzymes involved (2, 4). The enzymes required to synthesize a HS chain have all been cloned. Notably, several of these proteins, including species committed to polymer modifications, occur in multiple forms that are encoded by different genes. Some of these species have been shown to differ with regard to kinetic properties and/or substrate specificity from the respective homologous forms (7–10).

The IdoUA units of heparin and HS chains occur in widely different structural contexts, which range from the minimally sulfated -GlcNSO₃-IdoUA-GlcNAc_n-sequence found in HS molecules to the extensively sulfated -GlcNSO₃(6-O SO_3^-)-IdoUA(2-O SO_3^-)-GlcNSO₃(6-O SO_3^-)- structure typical of heparin (4). These findings raise the question as to whether the CS-epimerase, similar to other enzymes in the same biosynthetic

electrophoresis, kb, kilobase pair; PCS, fetal calf serum IgG, base pair; ORF, open reading frame; PCR, polymerase chain reaction; RT-PCR, reverse transcriptase-PCR; NDST, N -deacetylase/ N -sulfotransferase.

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